

# Amplifying Twist Multiplexed Gene Fragments

**For Research Use Only (RUO). Not for use in diagnostic procedures.**

Twist Multiplexed Gene Fragments (MGF) are diverse collections of double-stranded gene fragments synthesized using our silicon-based DNA writing technology. Our synthesis platform enables massively parallel production of hundreds of thousands of high-quality, accurate fragments per run. Multiplexed Gene Fragment sequences are available from 301-500 nucleotides and pool sizes start at 1,000 sequences with no

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## QUESTIONS?

Get in touch at [sales@twistbioscience.com](mailto:sales@twistbioscience.com) or learn more at

Twist uses phosphoramidite chemistry for synthesis. Compared to conventional column-based synthesizers, our synthesis platform miniaturizes the synthesis process, which reduces reaction volumes and increases throughput. PCR amplification on the initial synthesis material is very challenging; to mitigate the risk of poor amplification results, Twist is performing this step for you. If you need more DNA mass for cloning, are amplifying out subpools, or are adding a tailed primer to this

## RECOMMENDED PCR AMPLIFICATION PROTOCOL

The protocol below offers a starting point for PCR amplification. Twist Multiplexed Gene Fragments are delivered with Gs and GvS

**1** Prepare a stock solution of your Multiplexed Gene Fragments by resuspending in 10 mM Tris buffer, pH 8.0 to any concentration. A final concentration of 0.5 ng/ $\mu$ l is needed for the PCR Reaction.  
Stock solution concentration (ng/ $\mu$ l) = Total yield (ng) / resuspension volume ( $\mu$ l)

**2** Use the KAPA HiFi HotStart PCR Kit (Catalog #KK2502) to perform PCR.

### PCR REACTION COMPONENTS

| COMPONENT   | FINAL CONCENTRATION | PER 25 $\mu$ L REACTION |
|---|---------------------|-------------------------|
| 2X KAPA HiFi HotStart ReadyMix (contains dNTP and Polymerase)                   | 1x                  | 12.5 $\mu$ l            |
| 10 $\mu$ M Forward Primer   | 0.3 $\mu$ M         | 0.5 $\mu$ l             |
| 10 $\mu$ M Reverse primer   | 0.3 $\mu$ M         | 0.5 $\mu$ l             |
| Twist Multiplexed Gene Fragments (template input should be no more than 0.5 ng) | 0.5 ng/ $\mu$ l     | 1.0 $\mu$ l             |
| PCR grade water   | —                   | 10.5 $\mu$ l            |

### PCR REACTION CONDITIONS

| CYCLING STEP                         | TEMPERATURE                            | DURATION |
|--------------------------------------|--|----------|
| <b>1</b> Initialization Denaturation | 2 min at 95°C                          | 1x       |
| <b>2</b> Denaturation                | 20 sec at 98°C                         | 8 Cycles |
| <b>3</b> Annealing                   | 15 sec at 60°C or optimum temperature* |          |
| <b>4</b> Extension                   | 1 min at 72°C                          |          |
| <b>5</b> Final Extension             | 2 min at 72°C                          | 1x       |
| <b>6</b> Hold                        | Hold at 4°C                            | 1x       |

\*Annealing temperature depends on primer sequences and must be optimized accordingly.

**3** Purify the PCR reactions with SPRI magnetic beads using a low bead-to-DNA ratio (0.8X).

## QUALITY ANALYSIS AND TROUBLESHOOTING

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1. I tried amplifying the pool using the recommended guidelines and my pool did not amplify.